

Listing of the Claims:

This Listing replaces all the prior versions and listings of claims.

1. (Cancelled)
2. (Currently amended) A method of amplifying a hairpin structure comprising converting a double stranded nucleic acid into the hairpin structure, wherein the double stranded nucleic acid contains at least one sequence of interest, and is referred to as a template nucleic acid, and wherein the template nucleic acid has an upper strand with a 5' and a 3' end and a lower strand with a 5' and a 3' end, the method comprising:
  - (a) ligating a first single stranded nucleic acid to the 5' end of the upper strand of the template nucleic acid, [[and]]
  - (b) ligating a second single stranded nucleic acid, which is non-complementary to the first single stranded nucleic acid to the 3' end of the lower strand of the nucleic acid,
  - (c) ligating a cap of single stranded nucleic acid to both the 5' end of the lower strand and the 3' end of the upper strand of the template nucleic acid, such that the 3' end of the upper strand and the 5' end of the lower strand are contiguous, thereby creating the hairpin structure;and further comprising performing polymerase chain reaction with a first primer that binds to at least a portion of the upper single stranded non-complementary region at the 5' end of the upper strand, and a second primer that binds to at least a portion of the lower single stranded non-complementary region at the 3' end of the lower strand.
- 3-5. (Cancelled)
6. (Currently amended) A method of amplifying a nucleic acid sequence of interest that generates a PCR-amplified product which is substantially free of polymerase-induced errors, comprising:
  - (a) providing a sequence of interest comprising a double stranded nucleic acid, referred to as a template nucleic acid, wherein the template nucleic acid has an upper strand with a 5' and a 3' end and a lower strand with a 5' and a 3' end,
  - [[a)]] (b) converting the nucleic acid sequence of interest into a first hairpin DNA structure, ~~wherein the sequence of interest comprises a double stranded nucleic acid, and is referred to as a template nucleic acid, and wherein the template nucleic acid has an~~

~~upper strand with a 5' and a 3' end and a lower strand with a 5' and a 3' end~~, by ligating a first single stranded nucleic acid to the 5' end of the upper strand of the template nucleic acid, ~~[[and]]~~ ligating a second single stranded nucleic acid, which is non-complementary to the first single stranded nucleic acid to the 3' end of the lower strand of the nucleic acid ~~[[;]]~~, and ~~further comprising~~ ligating a cap of a single stranded nucleic acid to both the 5' end of the lower strand and the 3' end of the upper strand of the template nucleic acid, such that the 3' end of the upper strand and the 5' end of the lower strand are contiguous, thereby creating the first hairpin structure;

~~[[b)]]~~ (c) amplifying the first hairpin DNA structure using PCR with a first primer that binds to at least a portion of the first single stranded nucleic acid, and a second primer that binds to at least a portion of the second single stranded nucleic acid to produce a plurality of linear double stranded PCR products, wherein the double stranded PCR product comprises an amplified sequence of interest and its complementary sequence flanked 5' and 3' by the first and the second single-stranded nucleic acid sequences;

~~[[c)]]~~ (d) converting the linear double stranded PCR products into a plurality of second hairpin structures ~~by converting the linear double stranded PCR products into a plurality of second hairpin structures~~ by a method which induces ~~[[d]]~~ denaturation of the linear double stranded PCR products into single stranded PCR products, followed by sudden renaturation, wherein the amplified sequence of interest and its complement within each single strand hybridize during renaturation, thereby forming a hairpin structure;

~~[[d)]]~~ (e) identifying from the second hairpin structures mismatch containing hairpin structures that comprise gaps in binding between the sequence of interest and its complementary sequence in the double-stranded region of the second hairpin structure wherein the gaps are a result of polymerase-generated nucleotide changes, insertions, or deletions, and

~~[[e)]]~~ (f) removing such mismatch containing hairpin structure, and collecting the DNA that contains no mismatches.

7. (Currently amended) The method of claim 6, wherein the ~~[[a]]~~ method which induces denaturation followed by sudden renaturation is selected from the group consisting of (a) heat denaturation followed by rapid cooling, (b) addition of sodium hydroxide followed

- by sudden neutralization of the solution, and (c) addition of formamide followed by sudden removal of formamide.
8. (Previously presented) The method of claim 6, wherein the mismatch containing hairpin structures that contain PCR-induced errors and that have a mismatch in the double stranded region are separated from hairpin DNAs which do not contain mismatches by a method which recognizes DNA containing a mismatch.
  9. (Original) The method of claim 8, wherein the method which recognizes DNA containing mismatches is selected from the group consisting of dHPLC-mediated fraction collection, denaturing gradient gel electrophoresis (DGGE), constant denaturant gel electrophoresis (CDGE), constant denaturant capillary electrophoresis (CDCE), and an enzymatic-based separation method.
  10. (Original) The method of claim 9, wherein the enzymatic-based separation method is performed either in solution or bound to a solid support, and the enzyme is at least one enzyme selected from the group consisting of mismatch-recognition enzymes MutS, MutY, and TDG; Cel I; resolvases; endonuclease V; cleavases, and exonucleases.
  11. (Cancelled)
  12. (Cancelled)
  - 13.-16. (Cancelled)
  17. (Cancelled)
  - 18-35. (Cancelled)
  36. (Previously presented) The method of claim 6, wherein concentration of the first and the second primer in step (b) is equal to each other (~~“regular” PCR~~).
  37. (Previously presented) The method of claim 6, wherein concentration of the first and the second primer in step (b) is unbalanced (~~“asymmetric” PCR~~).
  38. (Previously presented) The method of claim 8 further comprising an assay consisting of mutation detection, mutation analysis, polymorphism detection, polymorphism analysis, microsatellite analysis, cloning, and protein functional analysis of the separated hairpin DNAs which do not contain mismatches.

39. (New) The method of claim 38, wherein the method of mutation or polymorphism detection is selected from the group consisting of PCR, PCR/RE/LCR, MutEx-ACB-PCR, RFLP analysis, and APRIL-ATM.